

A Comparison of Conventional, Revascularized and Bioengineered Methods of Recurrent Laryngeal Nerve Reconstruction Jordan P. Sand, MD¹; Andrea M. Park, MD¹; Neel Bhatt, MD¹; Shaun C. Desai, MD¹; Laura Marquardt, MS²; Shelly Sakiyama-Elbert, PhD²; Randal C. Paniello, MD, PhD¹ ¹Department of Otolaryngology-Head and Neck Surgery, Washington University School of Medicine ²Department of Biomedical Engineering, Washington University

ABSTRACT

Importance:

Damage to the recurrent laryngeal nerve (RLN) is highly detrimental to voice, swallow and cough. Nerve sacrifice is often necessary for thyroid or other cervical malignancies. The optimal reconstitution of this nerve after injury is unknown.

Materials and Methods:

An established canine model of RLN injury was utilized. Laryngeal adductor pressures (LAPs) were measured with a pressure transducer

INTRODUCTION

Although there is some inherent capability for nerves to regenerate, recovery after damage to a peripheral nerve is frequently unpredictable and can result in a poor outcome. When a nerve is completely transected, the distal and proximal stump can often be directly repaired via microsurgical suturing. However, in many cases this primary nerve repair may not be feasible and a bridging component of tissue is used to provide a tension free anastamosis (Siemionow 2009). Situations where this typically occurs are after trauma or during a cancer operation. When selecting a graft, surgeons usually choose to utilize an autologous nerve graft harvested from the patient's body. The use of an autograft can be limited by size mismatch among the nerve tissues and also by a lack of sufficient donor sites (Burnett 2004, Schmidt 2003). Several of these types of grafts exist; including skeletonized cable grafts, acellular nerve graft substitutes or vascularized nerve grafts. When these nerves are repaired under tension, there is almost always a poor outcome (Terzis 1975, Millesi 1981). Other environmental and patient inherent factors can also influence the ultimate success of nerve grafting, including the diameter of the nerve graft (Lux 1986). Free cable grafts have been observed to have central necrosis that is thought to be detrimental to the regeneration of the advancing axons (Lux 1986). One proposed technique for avoiding central necrosis is to place the nerve in a well-vascularized bed (Lux 1986). This technique assumes that the vascularized bed is sufficient to sustain the never graft and ultimately minimal damage would occur. Within the head and neck, most sites are closely related to the large vessels of the neck, but sites of relative low blood flow exist, particularly around the trachea. A different technique to avoid necrosis involves the use of vascularized nerve grafts that contain the surrounding vessels in addition to the neural tissue. These vessels are anastomosed to nearby vasculature which can provide native blood flow to the graft and potetially better performance, as has been demonstrated in the upper extremites (Lux 1986, Terzis 2009). Other grafting techniques, including the use of silastic or polyglycolic acid conduits to help guide grafts, have also been proposed \with some success in treated peripheral nerve injuries (Isaacs 2014). Further loading of these conduits with nerve growth factors, including glial cell-derived neurotrophic factor and neurotrophin-3, has been

METHODS AND MATERIALS

An established canine model of RLN injury was utilized (Paniello 2001). In this model, canine subjects undergo anesthesia per standard protocol and a permanent tracheostomy is placed. RLNs are dissected free and laryngeal adductor pressures (LAPs) are measured with a pressure transducer balloon via electrical stimulation of the nerve. The LAPs are again measured 6 months post-injury as a primary outcome of reconstruction. The recurrent laryngeal nerve is then injured and reconstructed with one of the study methodologies listed in table 1. For the acellular, revascularized and conventional cable grafts, a 5 cm length of RLN was reconstructed. Revascularized nerve grafts were harvested from the saphenous nerve, artery and vein and microvascular anastamosis was performed to the cranial thyroid artery and internal jugular vein. Acellular nerve allografts (5 cm, n=6) were harvested from canine saphenous nerves, engineered via established protocols and implanted into research subjects. The nerve conduits were each 2 cm long and had 0.5 cm of the transected nerve endings loaded into each side, permitting a 1 cm gap. Conventional cable grafts (5 cm, n=6) were also harvested from the saphenous nerve...

DISCUSSION (CONT)

The use of acellular nerve grafts and neural conduits has grown in popularity as this technology improves (Isaacs 2014). A number of inherent disadvantages exist to using autologous nerve grafts, including the limited availability of a suitable nerve as well as donor site morbidity such as scarring and numbness. There are FDA approved commercially available acellular grafts and conduits in use for multiple indications today. This is the first study to compare the use of vascularized nerve grafts, acellular nerve grafts and specialized coupling techniques with nerve growth factors for recurrent laryngeal nerve repair. This study has revealed that these specialized methods do not result in additional benefit compared with simple cable grafting or primary repair for reconstruction of the recurrent laryngeal nerve.

balloon pre-injury and 6 months post-injury via electrical stimulation of the RLN. The recurrent laryngeal nerve was reconstructed after simple transection directly or with two types of nerve conduits (silastic and neurotube). The nerve conduits were each 2 cm long and had 0.5 cm of the transected nerve endings loaded into each side, which permitted a 1 cm gap. The silastic conduit reconstructed with an empty carrier (n=4), a fibrin-embedded carrier (n=4), a fibrinembedded glial cell-derived neurotrophic factor carrier (n=8) or a fibrin-embedded neurotrophin-3 carrier (n=8). The neurotube conduit utilized a woven polyglycolic acid construct (n=11). Values were compared with transection of the RLN and direct neurorrhaphy (n=16). For reconstruction of the RLN with an intervening graft, a 5 cm segment of RLN was removed. Revascularized nerve grafts (5 cm, n=4) were harvested from the saphenous nerve, artery and vein with microvascular anastamosis performed to the cranial thyroid artery and internal jugular vein. Conventional cable grafts (5 cm, n=6) were also harvested from the saphenous nerve. As a comparison group, a section of RLN was removed and anastomosed in reverse configuration (2 cm, n=8). Acellular nerve allografts (5 cm, n=6) were harvested from canine saphenous nerves, engineered via established protocols and implanted into

Table 1

ethods of Recurrent Laryngeal Nerve Repair (number in experimental group)
ansection with direct neurorrhaphy (n=16)
verse autograft (n=8)
nventional cable graft (n=6)
vascularized nerve graft (n=4)
ellular (bioengineered) nerve allograft (n=6)
astic conduit with empty carrier (n=4)
astic conduit with fibrin-embedded carrier (n=4)
astic conduit with fibrin-embedded glial cell-derived neurotrophic factor carrier (n=8)
astic conduit with fibrin-embedded neurotrophin-3 carrier (n=8)
eurotube conduit of woven polyglycolic acid (n=11)

RESULTS

Simple RLN transection with direct neurorrhaphy provided 55.5% (\pm 12.5%) recovery of baseline LAPs. Reverse autografts provided 60.8% (\pm 27.5%) recovery of the baseline LAPs. The revascularized grafts and conventional nerve grafts provided a range of recoveries of LAPs. All revascularized nerve grafts were noted to have patent blood supply at canine sacrifice. Two of eleven neurotube reconstructions provided a measurable LAP with an average recovery of 37.1% of baseline. The other 9 neurotube reconstruction did not provide any measureable benefit. Reconstruction with an acellular nerve graft or a neural conduit in any condition provided no measurable LAP recovery.

CONCLUSIONS

Conventional nerve grafting provided equivalent recovery of laryngeal adductor pressures following recurrent laryngeal nerve repair as to a simple repair or a reverse autograft. Revascularized nerve grafts did not appear to provide additional recovery benefit. The use of bioengineered acellular nerve grafts and nerve conduits for reconstruction resulted in poor recovery of recurrent laryngeal nerve function.

REFERENCES

- 1. Paniello RC, West SE, Lee P. Laryngeal reinnervation with the hypoglossal nerve. I. Physiology, histochemistry, electromyography, and retrograde labeling in a canine model. Ann Otol Rhinol Laryngol. 2001 Jun;110(6):532-42.
- 2. Terzis JK, Kostopoulos VK. Vascularized ulnar nerve graft: 151 reconstructions for posttraumatic brachial plexus palsy. Plast Reconstr Surg. 2009 Apr;123(4):1276-91.
- 3. Millesi, H. Reappraisal of nerve repair. Surg Clin North Am 1981;61(1):321.
- Terzis, J., Faibisoff, B., and Williams, B. The nerve gap: suture under tension vs. graft. Plast Reconstr Surg 1975;56(1):166.
- 5. Szynkaruk M, Kemp SW, Wood MD, Gordon T, Borschel GH. Experimental and clinical evidence for use of decellularized nerve allografts in peripheral nerve gap reconstruction. Tissue Eng Part B Rev. 2013 Feb;19(1):83-
- 6. Kimata Y, Sakuraba M, Hishinuma S, Ebihara S, Hayashi R, Asakage T. Free vascularized nerve grafting for immediate facial nerve reconstruction. Laryngoscope. 2005 Feb;115(2):331-6.
- 7. Yoo YM, Lee IJ, Lim H, Kim JH, Park MC. Vein wrapping technique for nerve reconstruction in patients with thyroid cancer invading the recurrent laryngeal nerve. Arch Plast Surg. 2012 Jan;39(1):71-5.
- 8. Yumoto E, Sanuki T, Kumai Y. Immediate recurrent laryngeal nerve reconstruction and vocal outcome. Laryngoscope. 2006 Sep;116(9):1657-61.

research subjects.

Results:

Simple RLN transection with direct neurorrhaphy provided 55.5% (\pm 12.5%) recovery of baseline LAPs. Reverse autografts provided 60.8% (\pm 27.5%) recovery of the baseline LAPs. Revascularized and conventional nerve grafts provided a range of recoveries of LAPs. All revascularized nerve grafts were noted to have patent blood supply at canine sacrifice. Two of eleven neurotube reconstructions provided a measurable LAP with an average recovery of 37.1% of baseline. Reconstruction with an acellular nerve graft or a neural conduit in any condition provided no measurable LAP recovery.

Conclusions:

Conventional nerve grafting provided equivalent recovery of laryngeal adductor pressures following recurrent laryngeal nerve repair as to a simple repair or a reverse autograft. Revascularized nerve grafts did not appear to provide additional recovery benefit. The use of bioengineered acellular nerve grafts and nerve conduits for reconstruction resulted in poor recovery of recurrent laryngeal nerve function. attempted with demonstrated improved results in nerve regeneration in previous study (Pfister 2007, Moore 2010, Johnson 2009).

The recurrent laryngeal nerve (RLN) is sometimes sacrificed while undergoing thyroidectomy for malignancy. This nerve is responsible for the movement of the ipsilateral glottis, and its loss can be highly detrimental to a patient's ability to voice, swallow and cough. Numerous studies have been done with regard to primary repair of this nerve but occasionally there is a gap in the nerve that requires grafting (Chou 2003). The sacrifice of the nerve is often unplanned, and as such, repair of the nerve will need to be immediate to ensure the lowest amount of morbidity associated its loss. Techniques typically utilized by head and neck surgeons include using free ansa nerve cable grafts or great auricular nerve cable grafts (Li 2013). One group reported the use of free ansa grafts followed by vein wrapping and ultimately showed the there were better results than with conventional reconstruction (Yoo 2012). Further studies have showed that immediate reconstruction of the recurrent laryngeal nerve during surgery using grafts or

direct anastomosis showed reasonable voice outcomes (Yumoto 2006). While these methods function to a certain extent, they can certainly be improved upon by evaluating other known methods of nerve repair utilized in other situations. This study searches to evaluate nerve function after repair with multiple methodologies to to assess for an optimal method of RLN reconstruction.



DISCUSSION

The use of vascularized nerve grafts, acellular nerve grafts, neural conduits or nerve growth facturs in the head and neck or for the RLN has been extremely limited. One group demonstrated that utilizing a free vascularized nerve graft for the facial nerve immediately following sectioning in a select group of high risk patients and demonstrated that muscle movement recovered satisfactorily (Kimata 2005). A further case reported use of a vascularized lateral femoral cutaneous nerve graft to reconstruct the facial nerve (Kashiwa 2010). Another report looked at using neural conduits for canine RLN reconstruction and showed regeneration with a polyglycolic acid tube (Kanemaru 2003).

- 9. Lux P, Breidenbach W, Firrell J. Determination of temporal changes in blood flow in vascularized and
- nonvascularized nerve grafts in the dog. Plast Reconstr Surg. 1988 Jul;82(1):133-44.
 10. Brooks DN, Weber RV, Chao JD, Rinker BD, Zoldos J, Robichaux MR, Ruggeri SB, Anderson KA, Bonatz EE, Wisotsky SM, Cho MS, Wilson C, Cooper EO, Ingari JV, Safa B, Parrett BM, Buncke GM. Processed nerve allografts for peripheral nerve reconstruction: a multicenter study of utilization and outcomes in sensory, mixed, and motor nerve reconstructions. Microsurgery. 2012 Jan;32(1):1-14.
- 11. Brenner MJ, Hess JR, Myckatyn TM, Hayashi A, Hunter DA, Mackinnon SE. Repair of motor nerve gaps with sensory nerve inhibits regeneration in rats. Laryngoscope. 2006 Sep;116(9):1685-92.
- 12. Kawamura DH, Johnson PJ, Moore AM, Magill CK, Hunter DA, Ray WZ, Tung TH, Mackinnon SE. Matching of motor-sensory modality in the rodent femoral nerve model shows no enhanced effect on peripheral nerve regeneration. Exp Neurol. 2010 Jun;223(2):496-504.
- 13. Chou FF, Su CY, Jeng SF, Hsu KL, Lu KY. Neurorrhaphy of the recurrent laryngeal nerve. J Am Coll Surg. 2003 Jul;197(1):52-7.
- 14. Li M, Liu F, Shi S, Chen S, Chen D, Zheng H. Bridging gaps between the recurrent laryngeal nerve and ansa cervicalis using autologous nerve grafts. J Voice. 2013 May;27(3):381-7.
- 15. Sanuki, T., Yumoto, E., Minoda, R., & Kodama, N. The role of immediate recurrent laryngeal nerve reconstruction for thyroid cancer surgery. Journal of Oncology, vol 2010, 7 pages, 2010.
- 16. Jesuraj NJ, Santosa KB, Macewan MR, Moore AM, Kasukurthi R, Ray WZ, Flagg ER, Hunter DA, Borschel GH, Johnson PJ, Mackinnon SE, Sakiyama-Elbert SE. Schwann cells seeded in acellular nerve grafts improve functional recovery. Muscle Nerve. 2014 Feb;49(2):267-76.
- 17. Gulati AK, Cole GP. Immunogenicity and regenerative potential of acellular nerve allografts to repair peripheral nerve in rats and rabbits. Acta Neurochir (Wien) 1994;126:158–164.
- 18. Fox IK, Jaramillo A, Hunter DA, Rickman SR, Mohanakumar T, Mackinnon SE. Prolonged cold-preservation of nerve allografts. Muscle Nerve 2005;31:59–69.
- 19. Siemionow M, Brzezicki G. Chapter 8: current techniques and concepts in peripheral nerve repair. Int Rev Neurobiol 2009;87:141–172.
- 20. Burnett MG, Zager EL. Pathophysiology of peripheral nerve injury: a brief review. Neurosurg Focus 2004;16:E1.
- 21. Schmidt CE, Leach JB. Neural tissue engineering: strategies for repair and regeneration. Annu Rev Biomed Eng 2003;5:293–347.
- 22. Isaacs J, Browne T. Overcoming short gaps in peripheral nerve repair: conduits and human acellular nerve allograft. Hand (N Y). 2014 Jun;9(2):131-7.
- 23. Pfister LA, Papaloïzos M, Merkle HP, Gander B. Nerve conduits and growth factor delivery in peripheral nerve repair. J Peripher Nerv Syst. 2007 Jun;12(2):65-82.
- 24. Moore AM, Wood MD, Chenard K, Hunter DA, Mackinnon SE, Sakiyama-Elbert SE, Borschel GH. Controlled delivery of glial cell line-derived neurotrophic factor enhances motor nerve regeneration. J Hand Surg Am. 2010 Dec;35(12):2008-17.
- Johnson PJ, Parker SR, Sakiyama-Elbert SE. Controlled release of neurotrophin-3 from fibrin-based tissue engineering scaffolds enhances neural fiber sprouting following subacute spinal cord injury. Biotechnol Bioeng. 2009 Dec 15;104(6):1207-14.
- Kanemaru S, Nakamura T, Omori K, Kojima H, Magrufov A, Hiratsuka Y, Ito J, Shimizu Y. Recurrent laryngeal nerve regeneration by tissue engineering. Ann Otol Rhinol Laryngol. 2003 Jun;112(6):492-8.





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Figure 1: Photo of a dissected canine hindlimb with saphenous nerve (white arrow) and it associated vasculature (black arrow).

Figure 2: Photo of dissected recurrent laryngeal nerve following removal of 5 cm of nerve and then placement of vascularized saphenous nerve graft showing two nerve anastomoses (green arrows), arterial anastomosis (white arrow) and vein anastomosis (blue arrow).