## THE RECONSTRUCTION OF SKULL BASE — HEALING AND PITFALL AND PEARLS DURING RECONSTRUCTION OF SKULL BASE

Jan Hemza, M.D., PhD., PhD., MPA, Dpt. of Neurosurgery, Faculty Hospital about saint Ann, Brno, Czech Republic

The healing process is linked to the process of inflammation, as described by Robert Wirchow (1821-1902).

The study about histological, biomechanical quality of dura mater on the skull base, quality of different part of skull base, together with lamina cribrosa. From this study proceeds the study of healling dura mater, including of skull base bone. The healing of skull base is affect with great numbers of factors – anatomical–embryonal, biomechanical, biophysical, chemical and biological. The work goals are determination of the dural quality of the skull base area under localisation and there are into anterior skull base fossa, which the most connected with paranasal spaces, the study of the most sensitivity area - lamina cribrosa. Author of study reported in your works different type of reconstruction, different material from reconstruction and comments on 32 yy. Of experiences with those problems.

The border is formed by the chondrogenic bone and especially the dura mater, which is histologically formed mainly by collagen, a mixture of different types. The most important boundary between neurocranium and splanchnocranium is the dura mater. The problem of healing is based on this basis.

The ossification of the skull is important for healing too. The skull forms from two parts: desmogenic and chondrogenic. Desmogenic is skull vault and chondrogenic is skull base. The way of embryology development of skull have influence to healing. The chondrogenic part of skull have during healing problem (pure healing) with interposition of tissue (dura, brain, arterficial material etc.) into bone defect.

In order for healing to be as effective as possible, it is advisable to respect the basic principles of physical, chemical and biological.

Physiological function of skull base	The healing under mechanic stress		The collagens are piezoelectric—left is topography of collagen molecul, right is piezoresponse		lecul,
<ul> <li>Static function :         <ul> <li>boder between neuro- and splachnocranium</li> <li>protection of brain, vessels, nerves</li> <li>fixation and sealing of vessels, nerves during penetration through skull base</li> </ul> </li> </ul>	the intermitent compression or shear stress	the enchondral ossification	Topography	Piezoresponse	

## Dynamic function :

- Biomechanic
- Hemodynamic
- Liquorodynamic
- Lymphodynamic
- Imunne and immunodynamic
- Electromagnetic continnum electrophysiology
- Control homeodynamic function
- Note: excepting hypophyseos functions.

## The type of collagen

- **The fibrillar collage** group I,II,IV,V (90% of collagen into organism II chondroid tissue, III embryonic exchanges to I during maturation, IV- basal membrane, V wall of vessels)
- **The mesh collagen** group IV,VIII,X C-NC domen component of suprastructures
- FACIT (fibril-associated collagen with interrupted triple helix) group IX, XII, XIV, XVI, XIX, XX, XXI a XXII (collagen IX 3 α-chains, all of any only 1 α-chain) shotter C-NC domen only 30-75 residuum, fibrillar collagen 260 reziduum)
- **The transmembrane collagen** group XIII, XVII, XXIII, XXV nad any collagen similer proteins macrophag's receptor MARCO, α-chain N-NC domen 3 subdomen intracellular, transmemebrane, extracellular, the extracelluler connecting domen α- helical coiled-coil structure
- **Any collagens** group XV,XVIII, XXVI, XXVIII only 1 α-chain with N-NC and C-NC domen **Triple helix** model of crystalic structure Pro-Pro-Gly 103



the traction stress	the intramembranous ossification
the constant compression stress (hydrostatic pressure)	the cartilago inhibit enchodral ossification
the high shear stress	the formation of fibrous tissue

The histology of dura mater—the colorish van Giesen on collagen





Improvement formation and adhesivity of cells (pulsing mechanical stress)

		Pitfall	Pearl	
OP		minimalized study for localisation of defects	precise study of defects localisation – CT, HRCT, MRI, MRI-cisternography, CT-cisternography, endoscopic diagnosis	
		only basic biochemical study	bacterial, imnunological and biochemical energy study with study of proteinemie and albuminemia	
		macrosurgery	presice revision of skull base – microscopically, endoscopically	
			remove the interposits	
		without removed mucosa	paranasal mucose totally removed in operative field, in frontal sinus closed frontonasal duct, cranialization sinuses	
		one layer reconstruction	multilayer reconstruction	
		non-vascularized material	minimally one vascularized layer (periosteal, galeal, omental, musculofascial, musculofascioasubcutaneous)	
		free placed material	suture layers and tissue glue	
	Osseous recontruction	into paranasal sinuses	autotransplant, hydroxyapatit, titanium mash, monomer, ioceramic, microplates, methylmethacrylate - no into paranasal sinuses, only above skull base toward intracranially	
			local antibiotical sponge course to paranasal sinuses	
. ОР			antiobiotic under sensitivity of bacterial flora and local antibiotics	
		when used lumbal drainage, but without regulation of pressure	intracranial pressure normal – non-high, non-lower; when used lumbal drainage with regulation of pressure in normal range (mechanical effect fot healing)	
		horizontal position, vertical early position	postoperative body position – 30° upper part, vertisalisation after more than 5-7 days, but step by step	
	Dietary	Hypoproteinemia, hypalbuminemia, catabolic state, immune disorder, collagen deficit	Hypernutrition, sufficient collagen in the diet and dietary supplements - hydrolysed collagen type I, III and curcumin, vitamin C, vitamin D3, amino acids glycine, proline and hydroxyproline, zinc, copper and sulphur Food: fish, orange, lemon, grapefruit and lime, eggs – proline, zinc sulphur, strawberries, raspberries, blueberries, cranberries or other tasty berries, bone broth – consommé, beans – collagen-forming amino acids and copper, poultry with skin, meat, crustaceans and molluscs – collagen-forming amino acids and zinc and copper peptid thymosin beta	

## **Types of skull base reconstruction:**

Endoscopic technique Transcranial technique - extradural - intradural

Basic surgery technic : "toast" technic and "cuff" technic

Author study this problem under this factors (anatomical-embryonal, biomechanical, biophysical, chemical and biological) long time and ruminate over individual influences on healing of skull base on foundation personal experiences 32yy. with reconstructions of skull base after trauma and the others skull base surgeries. He anatomizes those factors on basis of anatomical-embryonal, biomechanical, biophysical, biological studies too.

In this study there is advented on biomechanical nad biophysical quality of collagen and those influences on the healing (piesoelectric, pieromagnetic effect etc.).

The goal of study is refered to pittfalls et pearls during surgery reconstructions of skull base a to influences, which have be invoved in quality of healing of skull base. He specify some methods from biomechanical a physiological study.

The conclusion of the healing study is shown in the table above.

References:
1. F Boccafoschi, M Bosetti, P.M Sandra, M Leigheb, and M Cannes: Effects of mechanical stress on cell adhesion A possible mechanism for morphological changes, Cell Adh Migr. 2010 Jan-Mar; 4(1): 19–25.
2. Edmund Y.S. Chao and Nozomu Inoue: Biophysical stimulation of bone fracture repair, regeneration and remodelling, European Cells and Material vol 6. 2003, pp. 72-85
3. James J. Tomasek, Giulio Gabbiani ,Boris Hinz, Christine Chaponnier and Robert A. Brown: Myofibroblasts and mechanoregulation of connective tissue remodelling, nature Review, moleculer Cell Biology, Vol.3, May 2002, pp.349-363
4. Wang, Hai-Qiang MD; Li, Ming-quan MD; Wu, Zi-xiang MD; Zhao, Li MD, PhD: The Deep Fascia in Response to Leg Lengthening With Particular Reference to the Tension-Stress Principle, Journal of Pediatric Orthopaedics: January/February 2007 - Volume 27 - Issue 1 - pp 41-45
5. Wang HQ, Zhao L.: Impact of mechanical stress and tension-stress on angiogenesis in wound healing, Chin J Traumatol., 2006 Apr;9(2):118-24
6. J.Hemza: Controversies in the Management of open Fracture of the Skull Base; An Interdisciplinary Approach, Vol.19 May 2009, suppl.1, p.77
7. J. Hemza : The bridging vein of brain and movement of the brain – the anatomical, biomaterial and biomechanical study, The 1st IMACS International Conference on Computational Biomechanics and Biology, ICCBB 2007, September 10-13, 2007, Plzeň Czech Republic

8. J.Hemza, R. Sedlacek: The dura mater – the basic intracranial border?, The 9th Asio-Oceanian International Congress on Skull Base surgery, the 10th Symposium of KSHNS, Paradise Hotel Busan, Busan, November 2008, South Korea

9. J.Hemza: The biomechanical problems of thinwall brain veins, <u>Dissertation's thesis</u>, Charles University, Praque 2010

10. Melvin, J.W., McElhaney, J.H., Roberts, V.L., 1970. Development of a Mechanical Model of the Human Head - Determination of Tissue Properties and Synthetic Substitute Materials, 14th Stapp Car Crash Conference. Society of Automotive Engineers, p.SAE Paper No. 700903.

**11.** C.V.Gisolfi,F. Mora: The hot brain, survival, temperature, and the human body, The MIT Press, 2000

12. Antila S, Karaman S, Nurmi H, Airavaara M, Voutilainen MH, Mathivet T, Chilov D, Li Z, Koppinen T, Park JH, Fang S, Aspelund A, Saarma M, Eichman A, Thoams JL, Alitalo K..: Development and plasticity of meningeal lymphatic vessels, J Exp Med 2017 Dec 4;214(12):3645-3667. doi: 10.1084/jem.20170391. Epub 2017 Nov 15.

13. Louveau A, Smirnov I, Keyes TJ, et al. Structural and functional features of central nervous system lymphatics. *Nature*. 2015;523(7560):337-341. doi:10.1038/nature14432.

14. Aspelund A, Antila S, Proulx ST, et al. A dural lymphatic vascular system that drains brain interstitial fluid and macromolecules. *The Journal of Experimental Medicine*. 2015;212(7):991-999. doi:10.1084/jem.20142290.

15. Iliff JJ, Wang M, Liao Y, et al. A Paravascular Pathway Facilitates CSF Flow Through the Brain Parenchyma and the Clearance of Interstitial Solutes, Including Amyloid β. Science translational medicine. 2012;4(147):147ra111. doi:10.1126/scitranslmed.3003748.

16. Aspelund A, Tammela T, Antila S, et al. The Schlemm's canal is a VEGF-C/VEGFR-3–responsive lymphatic-like vessel. *The Journal of Clinical Investigation*. 2014;124(9):3975-3986. doi:10.1172/JCI75395.

17. McIlvried LA, Cruz JA, Borghesi LA, Gold MS. Sex-, stress-, and sympathetic post-ganglionic-dependent changes in identity and proportions of immune cells in the dura. *Cephalalgia : an international journal of headache*. March 2016:0333102416637832. doi:10.1177/0333102416637832.

18. Absinta M, Ha SK, Nair G, Sati P, Luciano NJ, Palisoc M, Louveau A, Zaghloul KA, Pittaluga S, Kipnis J, Reich DS. :Human and non humand primater meninges harbor lymphatic vessels that can be visualized noninvasively by MRI, *Elife*. 2017 Oct 3;6

19. V Lecco: Di una probabile modificazione delle fissure linfatiche della della parte dei seni venosi della dura madre, Arch Ital Otol Rinol Laringol, 1953

20. Li J., Zhou J., Shi Y.: Scanning electron microscopy of human cerebral meningeal stomata. Ann Anat 1996 Jun;178(3):259-61.

21. Moeller, M.N.: Intracranial pressure revisited: A novel intravascular receptor detected, Dp.t of ORL, Head and Neck Surgery University Copenhagen, Denmark, WSB 2016, Osaka Japan

22. Antoine Louveau, Igor Smirnov, Timothy J. Keyes, Jacob D. Eccles, Sherin J. Rouhani, J. David Peske, Noel C. Derecki, David Castle, James W. Mandell, Kevin S. Lee, Tajie H. Harris, Jonathan Kipnis : Structural and functional features of central nervous system lymphatic vessels, Nature. 523: 337–41, 2015

23. Aleksanteri Aspelund, Salli Antila, Steven T. Proulx, Tine Veronica Karlsen, Sinem Karaman, Michael Detmar, Helge Wiig, Kari Alitalo : A dural lymphatic vascular system that drains brain interstitial fluid and macromolecules. The Journal of Experimental Medicine. 212: 991–9, 2015

24. Iliff JJ, Wang M, Liao Y, Plogg BA, Peng W, Gundersen GA, Benveniste H, Vates GE, Deane R, Goldman SA, Nagelhus EA, Nedergaard M : A Paravescular Pathway Facilitates CSF Flow Throught the Brain Parenchyma and the Clearence of Interstitial Solutes, Icluding Amyloid β. Sci Trans Med. 4 (147): 147ra111, 2012.

25. Cserr HF, Harling-Berg CJ, Knopf PM:Drainage of brain extracellular fluid into blood and deep cervical lymph and its immunological significance. Brain Pathol. 2 (4): 269–76, 1992.

26. Antoine Louveau, Jonathan Kipnis : Dissection and immunostaining of mouse whole-mout meninges. Protocol Exchange, 2015

Jan Hemza M.D. PhD.,PhD.,MP.

E-mail: jan.hemza@fnusa.cz